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| **BLOOD CULTURE TIP SHEET** | |
| * Remove caps from culture bottles and disinfect culture bottles green tube top with alcohol pad. * Optimal blood draw volume is 20-22cc. 10cc per culture bottle.   \*\*\*If unable to obtain optimal blood volume, fill (**in this order**): aerobic (**blue**) bottle completely (3 mL minimum), then remainder, if any, in anaerobic (**purple**) bottle.   * Make sure to put site on label. **Do not cover bar code label on bottles.** * **Do not cover the window with the measuring device on the side (Lab needs to be able to ensure proper blood amounts are in the bottles.** * **Place labels VERTICALLY instead of around the bottle (Lab needs to be able to scan the barcode).**      * If using the pneumatic tube system, place green top in a separate bag towards the necks of the bottles. Place Each bottle in a separate sealed zip lock bag and then securely double bag the set. **Do not tube more than one set per tube** | |
| PERIPHERAL STICKS ONLY w/NO CENTRAL LINE | CENTRAL LINE + PERIPHERAL STICK  OR   1. CENTRAL LINE DRAWS |
| * **Green tops not needed** * Scrub proposed site of venipuncture with ChloraPrep™ for 30 seconds using frictional back and forth motion. Allow to air dry naturally for 30 seconds prior to venipuncture. * **DO NOT re-palpate prepared site due to introducing contamination into the culture.** * Cultures obtained from the same site must be at least 10 minutes apart; from different sites they may be obtained at the same time. * **\*PRINTING LABELS – 3 labels print out for each culture set whether or not utilizing green tops**   + Pick 2 labels from the 1st three that print, discard the third label: label purple & blue bottles.   + Pick 2 labels from the 2nd three that print, discard the third label: label the purple & blue bottles. * **Ensure each culture set has matching numbers (19RL #)** | * **One Green top per site (1 for peripheral and 1 for central)** * Remove current central line cap from line. Scrub the open port with alcohol. Attach **new cap** to lumen using proper sterile technique. * Disinfect new central line cap with alcohol for 30 seconds. Wait 30 seconds for alcohol to dry. * Attach 10-20cc luer lock syringe to new cap. Aspirate blood filling whole syringe. **DO NOT FLUSH line prior to attaching syringe and do not waste any blood.** * Attach red access transfer device to blood-filled syringe and transfer blood. Allow blood to flow passively into each bottle. **Do not force blood into bottles!** * Wait at least 10 minutes before obtaining second cultures set from the same line. Make sure to print new stickers at timing of second draw. * **Ensure each culture set has matching numbers (19RL #)** |

