

AHRO Laboratories' Catalog of Tests U-Z

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Approvals

- Signature: Karen E Corson, Director, Laboratory signed on 7/22/2020, 2:43:05 PM
 - Signature: Alexandra Reichman, Physician signed on 7/22/2020, 4:13:20 PM
 - Signature: Karen E Corson, Director, Laboratory signed on 4/26/2022, 4:24:25 PM
 - Signature: Alexandra Reichman, Physician signed on 4/28/2022, 9:35:58 AM
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Subject/Title: **AHRO Laboratories Catalog of Tests U - Z**

Review the following alphabetical listing of our lab test catalog for each of our offered tests. Each page has at least the following elements:

- Test Name**
- Alternate Test Name**
- LIS/HIS Code**
- Storage/Transportation**
- Specimen Requirements**
- Optimal Volume**
- Minimal Volume**
- Post Testing Storage**
- Patient Preparation**
- Collection Instructions**
- Reference Ranges**
- Methodology**
- Additional Information**

AHRO Laboratories Catalog of Tests U - Z

Uric Acid (Blood or Urine)	
Alternate Test Name	
HIS Code / LIS Code	Blood: URIC Urine, Random: URICR
Where Performed	AHRO
Transportation/Storage	Blood: 3-5 days at 2-8°C Urine: 24 hours at 2-8°C
Specimen Requirements	Heparinized Plasma or Serum or Urine
Optimal Volume	1 mL
Minimal Volume	0.25 mL
Patient Preparation	None
Reference Ranges	Blood: Male: 2.6 – 8.7 mg/dL Female: 2.6 – 8.0 mg/dL Urine, Random: No Range
Methodology	Uricase
Additional Information	

Urinalysis	
Alternate Test Name	UA, Routine Urine
HIS Code / LIS Code	1234, UMAC
Where Performed	RMH
Transportation/Storage	N/A
Specimen Requirements	10 -12 ml of clean catch random urine
Optimal Volume	15 mL
Minimal Volume	2.0 mL
Storage	Room Temperature if test is to be performed within 2 hours, 2 - 8° C if delay is anticipated
Patient Preparation	None
Reference Ranges	See urinalysis report
Methodology	Bayer Multistix 10SG dipsticks and phase microscopy for manual microscopy, laser light scatter for automated microscopy.
Additional Information	Microscopic exam is done on those urines, which show positive nitrite, leukocyte esterase, blood or protein per good laboratory practice.

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Urine Culture	
Alternate Test Name	Urine C&S
HIS Code / LIS Code	CURINE
Where Performed	RMH
Transportation/Storage	Urine should be immediately transferred to a BD transport tube. Store and transport at room temperature
Preferred Specimen	Clean voided urine or urine collected via a catheter. The urine must be drawn into a BD transport tube.
Instructions	<ol style="list-style-type: none"> 1. Open bag and remove cup and towelettes 2. Unscrew cap of the cup. Place cap on counter with "straw" facing upward. TO AVOID CONTAMINATION, DO NOT TOUCH INSIDE OF CUP, CAP OR STRAW. 3. Cleanse yourself with towelettes as follows: 4. Male: Wipe head of penis in a single motion with one towelette. Repeat with remaining towelette. If not circumcised, hold foreskin back before cleansing. Urinate a small amount into toilet or bedpan. 5. Female: Separate the labia. Wipe inner labial folds front to back in a single motion with one towelette. Wipe down through center of labial folds with remaining towelette. Keep the labia separated and urinate a small amount into toilet or bedpan. 6. Place cup under stream and continue to urinate into cup and collect specimen. 7. Finish voiding into toilet or bedpan. 8. Replace cap on cup. Tighten cap securely. <p style="text-align: center;">CAUTION: SHARP NEEDLE LOCATED UNDER CAP LABEL. DO NOT REMOVE LABEL FROM CAP</p>
Laboratory instructions	<p><i>If using the Vacutainer collection cup with the needle sampler in the lid of the collection cup:</i></p> <ol style="list-style-type: none"> 1. Remove the yellow label on the lid of the cup. 2. Press the gray rubber septum of the provided gray cap tube firmly down onto the needle in the cap of the collection cup. 3. Allow to fill. 4. Remove the tube and shake vigorously. 5. Replace the yellow label on the cap after sampling. 6. Label and transport to RMH. Room temperature storage and transport. 7. <i>If using the straw transport kit:</i> Submerge tip of transfer device to bottom of urine container. 8. Place BD Vacutainer Tube in holder portion of transfer device. Push tube all the way into holder. 9. Hold in position until urine stops flowing in to tube. 10. Remove tube from transfer device and set aside. Lift transfer device and allow urine to drain from straw. Treat transfer device as a contaminated sharp and discard in a biohazard container approved for their disposal. 11. Shake tube vigorously. Label and send to RMH. Room temperature storage and transport.
Methodology	Conventional Culture
Reference Range	No colony forming units cultured
Set up Schedule	Processed daily. Preliminary Reports within 24 hours. Final report dependent upon growth.

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Urine Culture if indicated	
Alternate Test Name	
HIS Code / LIS Code	1234C, UACII
Where Performed	RMH
Transportation/Storage	N/A
Specimen Requirements	10 -12 mL clean catch, random urine
Optimal Volume	15.0 mL
Minimal Volume	2.0 mL
Storage	Room Temperature if test is to be performed within 2 hours, 2 - 8°C if delay is anticipated
Patient Preparation	None.
Reference Ranges	
Methodology	Bayer Multistix 10SG dipsticks and phase light microscopy for manual microscopy, Laser light scatter for automated microscopy.
Additional Information	Microscopic exam is done on those urines, which show positive nitrite, leukocyte esterase or protein. This test is to be used when the physician's order is "culture if indicated". When this test is ordered the laboratory will perform the routine urinalysis and if the Leukocyte esterase test or the nitrate test is positive or if in the microscopic exam >10 WBC's are seen per high power field, a culture will be ordered and set up on that specimen

Urine for Eosinophils	
Alternate Test Name	
HIS Code / LIS Code	1156, UAEOS
Where Performed	RMH
Transportation/Storage	Specimen should be run immediately after collection
Specimen Requirements	Freshly voided urine
Optimal Volume	10 ml of urine
Patient Preparation	None
Reference Ranges	<1 %
Methodology	Hansel Stain
Additional Information	Rare WBC's present in the sample could affect results since the test is reporting percentage of eosinophils found.

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Valproic Acid

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Alternate Test Name	Depakote
HIS Code / LIS Code	VALP
Where Performed	AHRO
Transportation/Storage	5 days at 2-8°C
Specimen Requirements	Heparinized Plasma or Serum
Optimal Volume	1 mL
Minimal Volume	0.25 mL
Patient Preparation	Draw –1-4 hours post oral dose if dose is known.
Reference Ranges	50 - 100 µg/mL
Methodology	Homogeneous Enzyme Immunoassay (EMIT)
Additional Information	Note last dose time and date with requisition in the LIS.

Vancomycin	
Alternate Test Name	
HIS Code / LIS Code	Random Level: VANCN Peak: VANCN Trough: VANCN
Where Performed	AHRO
Transportation/Storage	3 Days at 2-8°C
Specimen Requirements	Heparinized Plasma or Serum
Optimal Volume	1 mL
Minimal Volume	0.25 mL
Patient Preparation	Draw levels at: Peak: 30-60 minutes post I.V. dose Trough: Immediately before the next dose
Reference Ranges	Peak: 20.0 - 40.0 µg/mL Trough: 5.0 – 15.0 µg/mL
Methodology	Homogeneous Enzyme Immunoassay (EMIT)

VITAD

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Alternate Test Name	Total 25 OH Vitamin D
HIS Code / LIS Code	VITD25OH
Where Performed	RMH
Transportation/Storage	72 Hours at Room Temp / 7 Days at 2-8°C
Specimen Requirements	Green top Plasma, Serum, EDTA, Lithium-heparin, Sodium heparin
Optimal Volume	1 ml
Minimal Volume	0.5
Patient Preparation	None
Reference Ranges	30 – 100 ng/ml sufficiency, <20 ng/ml deficiency, > 100 toxicity.
Methodology	Two Step Competitive Binding immunoassay
Additional Information	No hemolysis

VIT B12 (Vitamin B12/Folate Battery – see below)	
Alternate Test Name	
HIS Code / LIS Code	VITB12
Where Performed	AHRO
Transportation/Storage	8 Hours at Room Temp / 24 Hours at 2-8°C
Specimen Requirements	Serum, alternate Heparin plasma, EDTA plasma
Optimal Volume	1.0 ml
Minimal Volume	0.5 ml
Patient Preparation	NONE
Reference Ranges	Normal 211 – 911 pg/ml, Deficient 32 – 246 pg/ml
Methodology	Direct Chemiluminescent technology
Additional Information	Avoid exposing sample to excess light