

# ADDRESSOGRAPH

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FEATHER RIVER HOSPITAL  
5974 PENTZ ROAD  
PARADISE, CA 95969  
(530) 877-9361

ORDERING DEPARTMENT(required)

PROCEDURE DATE (required)      ORDERING CLINICIAN First & Last name (required)

COPIES TO: \_\_\_\_\_



**PATHOLOGY SCIENCES MEDICAL GROUP**  
183 East 8th Avenue • Chico, CA 95926 • (530) 891-6244 • (800) 292-0760

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**CYTOLOGY REQUEST**

PATIENT NAME			DATE	
PREVIOUS NAME (if changed within last 5 yrs)			* DATE OF BIRTH *	
MAILING ADDRESS			AGE	SEX M F
CITY	STATE	ZIP	PHONE	

insurance info:  attached OR  completed below:

INSURANCE CO	
INSURANCE ID #	GROUP #
2 <sup>ND</sup> INSURANCE CO	
2 <sup>ND</sup> INSURANCE ID # AND GROUP #	

## Cytology

**CLINICAL HISTORY (Please do not enter the procedure itself, rather: state the REASON for the procedure):**

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**SPECIMEN TYPE and ANATOMIC SITE (required):**

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- Request AFB stain
- Request fungal/pneumocystis stain

**ANALYSIS ORDERED:**  Cytology

**FLOW CYTOMETRY SENT:**

Leukemia/Non-Hodgkin Lymphoma with DNA

OFFICE USE: RS

**Specimen Submission Guidelines – ALL specimens must be labeled with at least 2 patient identifiers**, placed in the plastic biohazard bag, and sealed. Requisition is placed in the exterior pocket of the plastic bag. Submit to the hospital lab and send with the next available courier to the PSMG Cytology Department.

**BODY CAVITY FLUIDS (Pleural, Pericardial and Peritoneal) DO NOT FIX**

Specimen < 1 Liter send entire specimen.

Specimen > 1 Liter mix specimens thoroughly and remove a 100mL aliquot.

- Send to the hospital lab as soon as possible.
- Refrigerate specimen if pick up is not within 24 hours.

**CSF (Cerebrospinal Fluid) DO NOT FIX Volume (3mL – 10mL)**

- In order to prevent cellular degeneration send cytology aliquot to hospital lab as soon as possible.
- Refrigerate specimen in the hospital lab until pick up, and send with the next available courier to the PSMG Cytology Department. Do not freeze specimen.

**PELVIC WASHING (Abdominal Wash) FIX**

- Send to the hospital lab as soon as possible.

**Sputum: FIX**

- Early morning, deep cough specimens are preferred. Instruct patient to rinse mouth with water prior to each collection.
- Have patient cough deeply from diaphragm and collect sputum.
- Mix specimen in original collection container with equal volume of Saccomanno fixative.

**Bronchial (or gastric) Washings: FIX**

- Mix specimen with equal volume of Saccomanno fixative.
- Submit to the hospital lab (if applicable) and send with the next available courier to the PSMG Cytology Department.

**Bronchoalveolar Lavage: FIX UNLESS...If requesting a lipid stain, do not add Saccomanno fixative & refrigerate**

- Submit to the hospital lab and send with the next available courier to the PSMG Cytology Department.

**BRUSHINGS (bronchial, gastric, other) FIX**

- Prepare and fix slides promptly.
- Label slides with patient's name and specimen type (in pencil).
- Rotate brush gently, but rapidly on clean glass slide.
- Fix slide immediately with spray fixative.
- Rinse the remaining material from brush or clip off the wire and place the entire brush in a container of Saccomanno fixative.

**URINE (Voided, Catheterized, Ileal Conduits) BLADDER WASHINGS FIX Minimum specimen volume is 25 mL**

- First morning urine should be avoided due to possible overnight cellular degeneration. A "clean catch"/ midstream specimen for women is recommended to avoid vaginal cell contamination.
- Hydrate the patient before collection.
- Mix entire specimen with equal volumes of Saccomanno fixative. If Saccomanno fixative is unavailable, refrigerate until pick up.
- Label the specimen container with specimen source details: voided, catheter, ileal, washing.

**CYST FLUID (Breast, Ovarian, Other) FIX**

- Mix specimen with equal volume of Saccomanno fixative.

**BREAST OR NIPPLE SECRETIONS FIX**

- Gently strip subareolar area and nipple with thumb and forefinger.
- When secretion occurs, allow a pea-sized drop to accumulate. Touch a clean slide to the nipple and withdraw slide quickly.
- Immediately spray slide with fixative.
- Repeat procedure until all secretions from the breast are collected on two or more slides.
- When dry, place slides in slide container.

**TZANCK/Viral/HSV/Herpes Preparation FIX**

- The Viral/Herpes/Tzanck Preparation is the scraping of a lesion or vesicle in order to determine possible viral origin.
- Write patient's name on the frosted end of 2 slides in pencil.
  - Slide One: Open a FRESH vesicle with a sterile blade, spatula or other appropriate instrument (you may also aspirate the lesion) and touch the instrument to the labeled slide.
  - Immediately spray fix with cytology (Pap) fixative. Please DO NOT AIR DRY the slide.
  - Slide Two: Scrape the vesicle and touch to the second slide. Do NOT smear.
  - Immediately spray fix with cytology (Pap) fixative. Please DO NOT AIR DRY the slide.
  - When dry, place slides into slide container.

**MISCELLANEOUS SMEARS (Skin Lesion, other) FIX**

- Label slides with patient's name and the source of the smear in pencil.
- Fix slides immediately with spray fixative.
- Allow to dry and place slide in slide carrier.

**FINE NEEDLE ASPIRATIONS (Breast, Head + Neck, Lung, other) Fixation is specimen dependent.**

- Telephone the Pathologist on call at 891-6244 for technical questions. For general questions ask for the Cytology Department at the same number. Label slides with the patient's name (in pencil).

**Needle note:** After specimen collection, needles can be rinsed in Saccomanno fixative. This rinsing, **but not the needle itself**, can then be safely sent to PSMG for processing if appropriate. The rinsed needle should be disposed of properly at clinic/hospital, and NOT sent on to PSMG.