

Specimen Collection and Transport Devices

Suitable containers are usually supplied by the hospital or stocked at nursing stations. Certain specialized or less commonly used transport containers are available only from the Microbiology lab. Refer to the pictures and instructions below for use of approved transport devices.

ESwab w/Regular flocked swab (White Cap)

When to Use: For all routine aerobic and anaerobic swab collections for bacterial and fungal culture (throat, nasal, ear, genital, superficial wound), Candida auris Screening NAAT, and Carbapenem Resistant Screening NAAT. Liquid media allows additional material for routine and fungus culture. If AFB culture is ordered, a separate ESwab collection is required due to volume requirements.



Description: System consists of clear liquid Amies medium in a flat-bottomed tube with a white cap. The outfit also contains a regular flocked swab. The uninoculated kits are stored at room temperature.

How to Use: Peel apart protective sleeve and take out tube. Remove flocked swab and use to collect sample. DO NOT POUR OUT LIQUID FROM TUBE! Carefully place swab into original tube, break off the swab shaft at colored breakpoint indicator mark. Recap the tube tightly.

ESwab w/Mini-tip flocked swab (Dark Blue Cap)

When to Use: For all routine and anaerobic swab collections of specimens from small orifices or difficult to reach areas such as nasopharyngeal, urethral, and conjunctival.



Description: System consists of clear liquid Amies medium in a flat-bottomed tube with a dark blue cap. The system also contains a mini-tip flocked swab. The uninoculated kits are stored at room temperature.

How to Use: Peel apart protective sleeve and take out tube. Remove flocked swab and use to collect sample. DO NOT POUR OUT LIQUID FROM TUBE! Carefully place swab into original tube, break off the swab shaft at colored scored indicator mark. Recap the tube tightly.

MSwab w/Regular flocked swab (Light Blue Cap)



When to Use: For MRSA NAAT and Staph aureus NAAT specimens only. NOTE: MSwab is NOT acceptable for specimens for routine culture, Candida auris NAAT, or Carbapenem Resistant Screen by NAAT.

Description: System consists of liquid molecular preservation medium in a flatbottomed tube with a light blue cap. The system also contains a regular flocked swab. The uninoculated kits are stored at room temperature.

How to Use: Peel apart protective sleeve and take out tube. Remove flocked swab and use to collect sample. DO NOT POUR OUT LIQUID FROM TUBE! Carefully place swab into original tube, break off the swab shaft at colored scored indicator mark. Recap the tube tightly.

Nares Specimen Collector – Saline w/Regular flocked swab (Various colored Caps)



When to Use: For SARS CoV-2 (COVID-19) NAAT specimens only.

Description: System consists of 3mLs of saline in various tube types. The system also includes either a regular flocked swab or a regular wound swab. The uninoculated kits are stored at room temperature.

How to Use: Open the package and take out tube. Remove swab and use to collect sample. Rotate the swab 5 times in each nostril. Be sure to sample from both nostrils. DO NOT POUR OUT LIQUID FROM TUBE! DO NOT PRE-MOISTEN THE SWAB!! Carefully place swab into original tube, break off the swab shaft at colored scored indicator mark. Recap the tube tightly.

Roche cobas[™] PCR Media

When to Use: For testing specimens collected for chlamydia, and gonorrhea testing, by molecular methods. Designed for transporting urine and female vaginal/endocervical swab, along with throat, rectal, and eye swab.

Description: Urine collection kit consists of a round-bottomed yellow-top tube with clear fluid and a sterile plastic pipette to be used for transferring patients' urine into the tube. Female swab collection kit consists of a round-bottomed vellow-top tube with clear fluid and 2 sterile swabs.

To Minimize the Risk of Invalid Test Results from Vaginal and Endocervical sources: Refrain from using carbomer-containing products to lubricate a speculum as part of the collection procedure or collect the sample prior to use of lubricant and/or performing pelvic exam. Ask patients to refrain from using feminine hygiene products for approximately 24 hours prior to the scheduled visit. If the patient is known to be using feminine hygiene products at time of collection, an alternative non-vaginal swab sample type may be considered, e.g., Urine.

How to Use:

Urine (Male and Female)

- Patient should not urinate 1 hour prior to collection.
- Instruct patient to collect first 10-50 mL of urine in sterile container (larger volumes decrease sensitivity of test, i.e., increase false negative results.)
- Use pipette to transfer urine into PCR Media tube, adding urine to bring the volume of sample to between the 2 urine fill lines as indicated by the arrows on the tube.
- Re-cap tube then invert X5.

Vaginal

- Insert ONE swab about 5 cm into vaginal opening. (Use woven swab.)
- Gently turn swab for 30 seconds against wall of vagina. Withdraw swab and immediately lower swab into tube until the dark line on shaft is aligned with the tube rim.
- Break off shaft at dark line and re-cap tube.
- Tubes received with 2 swabs will be rejected.

Endocervical

- Use first swab (woven swab) to clean excess mucus from cervical os (mucus • interferes with test and will result in an Invalid result.) DISCARD FIRST SWAB!
- Use second swab (flocked swab) to collect sample by inserting swab into ٠ endocervical canal and gently rotating 5 times in one direction.
- Lower swab into tube until the dark line on shaft is aligned with the tube rim.
- Break off shaft at dark line and re-cap tube.
- Tubes received with 2 swabs will be rejected.
- Urethral

No MALE collection kit for urethral samples; submit First voided Urine.



Urine Kit



Female Swab Kit

Universal Viral Transport Medium (UVT) with Flexible Flocked Mini-tip Swabs



When to Use: For collection and transportation of nasopharyngeal (NP) swabs for Influenza and Respiratory Syncytial Virus (RSV) rapid antigen testing, Influenza and RSV molecular testing; other Viral Respiratory Pathogen; and Bordetella molecular testing.

Description: System consists of a vial of medium (pink fluid with glass beads for macerating cells) in a flat-bottomed centrifuge tube with a red cap. The outfit also contains a single swab packet with a flexible flocked mini-tip swab. The uninoculated kits are stored at room temperature.

How to Use: Collect NP specimen, insert into the transport medium, break off shaft (leave swab in transport), tighten lid securely and refrigerate or freeze.

Stool Culture Transport (Para-Pak[®] C&S)



When to Use: Inpatient stool specimens should be collected in clean container and promptly delivered to laboratory within two hours. The pictured container is recommended for Outpatients as there may be a significant delay in transit. *Note:* Collection for Clostridium difficile toxin must be in a clean container without preservative. **Do not use this container.**

Description: Device consists of a single vial with an orange cap containing nonnutritive stool transport solution. A sample spoon is attached to the underside of the lid.

How to Use: Fill vial with stool to red line using a tongue depressor or the spoon attached to lid of vial. Cap tightly and mix or shake until stool is well emulsified.

Ova & Parasite (O&P) Transport (MCC[®] Total-Fix)



When to Use: All patient stool collections for Ova and Parasite examinations must be placed in this container.

Description: Consists of a single vial with a black cap containing a proprietary and ecological fixative (no mercury, formalin, or PVA) designed to preserve specimens for staining and concentration.

How to Use: Fill vial with stool to red line using a tongue depressor or the spoon attached to lid of vial. This will ensure the required three to one ratio of fixative to sample. Cap tightly and mix or shake until stool is well emulsified.

Sticky Paddle for Pinworm Collection



When to Use: For pinworm examination specimens.

Description: Individually packaged tubes contain a one-sided sticky paddle attached to the lid.

How to Use: Ideal collection time is early morning. Peel plastic wrap to remove tube and cap. Spread patient buttocks and apply sticky side of paddle to anal folds. Replace paddle in tube and twist to seal well. Label and deliver tube to laboratory.

Scabies Collection Kit



When to Use: For collection of skin scrapings for scabies examination.

Description: Each kit contains a tube containing 0.5 mL of mineral oil plus a sterile scalpel and pipet to collect and submerge the parasite-infested skin scrapings.

How to Use: Using the pipet, place drop of mineral oil from the tube on the sterile scalpel blade and allow oil to flow onto the papule. Scrape vigorously to remove the top of the papule (there should be flecks of blood in the oil.) Using the same pipet, transfer oil and scalpel material to test tube. Tap material down to submerge scrapings in remaining oil. Cap tube securely, label and deliver to laboratory.

Blood Culture Collection Vials (AFB Blood and Fungus Blood)



When to Use: The timing of blood cultures is a clinical decision. One bottle required for each order.

Description: Vials contain nutrient broth and lysing agents designed to optimize recovery of mycobacteria (AFB).

How to Use: Prior to injection of blood sample, remove colored cap from vial and prep septum with a 70% isopropyl alcohol wipe.

Venipuncture: Prep arm with ChloraPrep[®] antiseptic scrubbing gently with a back and forth motion for 30 seconds and allow to dry for 30 seconds.

Volume Recommendations:

3 to 5 mL optimum, 1 mL minimum volume into one Myco/F Lytic vial

If both AFB Blood and Fungus Blood Cultures are ordered, each order needs its own bottle drawn. They can be from the same venipuncture. Be sure to add optimal volume of blood to each bottle. When to Use: The timing of blood cultures is a clinical decision. Each set of blood cultures is a separate order and should be collected via a separate venipuncture.

Description: Vials contain nutrient broth with anticoagulant, resins or lysing additives designed to optimize growth of pathogens.

How to Use: Prior to injection with blood sample, remove colored cap(s) from vials and prep septa with 70% isopropyl alcohol wipes.

Venipuncture: Prep arm with ChloraPrep antiseptic scrubbing gently with a back and forth motion for 30 seconds and allow to dry for 30 seconds.

Volume Recommendations:

Adults:

16 to 20 mL blood distributed between two vials

- 8 to 10 mL into Aerobic vial
- 8 to 10 mL into Anaerobic Lytic vial

Difficult Adult and Older Pediatric Patients (> 6 yrs):

5 to 10 mL blood into Aerobic vial

3 mL to 5 mL blood into Aerobic vial – Label as suboptimal

Less than 3 mL blood - Specimen rejected; re-collect

Infant and Young Pediatric Patients:

1 to 3 mL into 1 vial into Peds vial

Less than 1 mL blood into one Peds vial – Label as suboptimal

Note: Collection of blood through intra-arterial or central venous lines is discouraged unless a concurrent set is collected peripherally.

Important: Multiple blood cultures collected from one venipuncture are considered **one** large-volume blood culture set.

Bone Marrow: Use 1.5 mL Sodium Heparin tube.



DO NOT USE FOR ADULTS OR OLDER PEDIATRICS (>6 YRS)

Ambient Temperature Transport System (ATTS) for Affirm Bacterial Vaginosis/Vaginitis DNA Panel



When to Use: For collection of vaginal specimens for Affirm[™] Bacterial Vaginosis/Vaginitis DNA Panel.

Description: Kit includes swab, stabilizing solution (dropper), transport tube and cap plus instructions.

How to Use:

- Open entire ATTS kit. Tear foil pouch, break ampoule by squeezing the sides of the plastic until the glass breaks. Express the contents into the provided sample tube. Dispose of glass ampoule in a safety container.
- Place patient in position for pelvic examination
- Insert speculum into vagina to permit visualization of posterior vaginal fornix
- Use sterile swab to obtain sample from posterior vaginal fornix. Roll swab against vaginal wall two to three times ensuring entire circumference of swab makes contact. Swab lateral vaginal wall while removing swab
- Place swab into vial containing stabilizing solution and break off shaft at scored area. Cap securely for transport to Laboratory.
- Follow appropriate labeling procedures at bedside.

Urine Culture for Microbiology Culture or Urinalysis

When to Use: For the collection of urine for microbiology culture or urinalysis.

Description: Urine Complete Cup Kit (3-part) -- Includes urine cup, two urine tubes, and soap towelette.

How to Use: The healthcare professional obtains the cup for the patient and cautions patient not to remove the cap label to protect against needlestick from the "sharp" contained in the integrated transfer device. If a kit is used, the healthcare professional should remove the tube(s) and place them in a protected location. If kits are provided to the patient, the patient should be directed to follow instructions on the bag for proper collection of a clean-voided, midstream urine specimen. Patient is instructed to give the urine specimen to the healthcare professional immediately after collection.

To transfer the specimen into evacuated tube(s):

- Place cup upright on clean, flat surface. Container may be tipped at an angle if specimen volume is limited.
- Peel back label on cap to expose the integrated transfer device.
- Place evacuated tube into cavity on cap, stopper down. Advance the tube over puncture point to pierce stopper. BD Vacutainer[®] C&S Preservative Urine Tubes should be filled first when collecting multiple tubes.
- Hold tube in position until filled.
- Remove tube from integrated transfer device.
- For all BD Vacutainer[®] Preservative Urine Tubes, mix tubes 8-10 times by inversion.
- Repeat steps if another tube is to be filled.
- Replace label over integrated transfer device cavity and reseal. Use caution to avoid contact with needle when replacing label.
- Label tube(s) or cup for transport to laboratory.
- Treat the screw cap of the cup as a contaminated sharp and discard in biohazard container approved for sharps disposal.

IMPORTANT NOTE: Do not use the UA tube (yellow/red top) for urine culture or it will be rejected when submitted to the lab for culture.







Unsuitable Transport for Microbiology Testing



Specimens submitted in CytoLyt (for cytological examination) or in formalin (for histologic testing) are inappropriate for culture because the preservatives affect the viability of microorganisms. Stains can be performed on the specimens to detect bacterial or fungal pathogens but the specimens will be rejected for culture.

NOTE: SPILLS IN TRANSIT AND IN THE TUBE SYSTEM AFFECT THE INTEGRITY OF SPECIMENS AND THE SAFETY OF YOUR CO-WORKERS!

DO NOT USE THIS CONTAINER FOR ANY LIQUID SPECIMENS!



USE THESE SCREW-CAP CONTAINERS

